Borderline–tuberculoid leprosy: clinical and immunological heterogeneity

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Summary The authors analysed some immunological criteria in leprosy patients diagnosed as borderline tuberculoid by the presentation of different grades of skin lesions as well as different grades of nerve involvement. Only 50% of the patients presented a single skin lesion and 58% had none or only one affected nerve. Nineteen patients (39.6%) showed a positive lepromin reaction (induration ≥ 5 mm).

Patients with a positive skin test had a greater number of skin lesions when compared with patients with a negative lepromin test. Fifty-seven percent of the patients were found to be positive using a lymphoproliferation test (LTT) in response to Mycobacterium leprae antigens. Positive LTT results did not correlate with the number of skin lesions, but patients unresponsive to LTT had a lesser extent of nerve involvement. Four out of 18 patients (22%) released high IFN-γ levels in PBMC culture stimulated by M. leprae. (mean U/ml ± SD = 142 ± 72). All of these 4 patients presented only one skin lesion, although three of them had more than one affected nerve.

Nineteen out of 21 patients (90.5%) showed no anti-PGL-1 antibodies in their serum. The low levels of anti-PGL-1 antibodies among these patients confirmed their tuberculoid background even in those with multiple skin lesions.

These findings seem to attribute an important role to IFN-γ in restraining the spreading of the infection in the skin, but IFN-γ may have an opposite effect on the nerves. The potential pathological effects of IFN-γ during the delayed type of hypersensitivity can be related to its ability to synergise with other inflammatory cytokines such as TNFα, IL-1β, and others.
Introduction

Leprosy is a chronic infectious disease that affects skin and peripheral nerves. The clinical spectrum in leprosy seems to correlate with the host immune response. T-cell-dependent immunity to *Mycobacterium leprae* is high in healthy-exposed individuals and in tuberculoid leprosy patients with localized disease, but is strikingly absent in lepromatous leprosy patients who have a high bacillary load and widely disseminated lesions. Thus, cell-mediated immunity (CMI) protects against dissemination of bacteria and disease. It has recently been hypothesized that the spectrum of leprosy reflects the balance between T-helper-cell populations (Th1 and Th2 CD4+ T cells). Indeed, analysis of mRNA expression in tissue samples of leprosy patients through RT-PCR showed two distinct lymphokine patterns. Tuberculoid lesions were characterized by prominent expression of IL-2 and IFNγ mRNA, a Th1 pattern associated with cell-mediated immunity. Moreover, lepromatous lesions primarily expressed IL-4, IL-5 and IL-10 mRNA, the Th2 pattern which favours humoral immunity and suppression of DTH.

IFNγ, a product of activated T lymphocytes, has been considered the major mediator at the effector end of the immune response. Its production correlates with the ability of mononuclear phagocytes to kill microorganisms. It was also demonstrated that household contacts of lepromatous patients who were unable to secrete IFNγ upon specific in vitro stimulation had a higher risk of developing leprosy as compared to high responders. Tuberculoid leprosy patients (TT, BT) are known to respond in vitro to *M. leprae* antigens, although nonresponder BT patients have also been reported. However, the in vitro immunological status of paucibacillary patients and its correlation to their clinical activity, lepromin response, and the occurrence of relapse has not yet been investigated. Nerve lesions and the extent of body area affected are important parameters in determining the treatment of leprosy patients. The aim of this study was to correlate the in vitro and in vivo immunological responses of tuberculoid leprosy patients with the extension of skin lesions and the involvement of peripheral nerves.

Material and methods

**PATIENT POPULATION**

Leprosy patients from the Leprosy Out-Patient Unit, Oswaldo Cruz Foundation, Rio de Janeiro, Brazil, were diagnosed according to the Ridley–Jopling classification. A total of 48 borderline–tuberculoid (BT) leprosy patients (18 male and 30 females), 15 to 78 years of age (mean ± sd = 41 ± 12), were studied. Patients were included in the World Health Organization (WHO) multidrug therapy (MDT) regime for paucibacillary patients. None of them had been previously treated. All of them received 6-monthly supervised doses of 600 mg rifampin and daily self-administered doses of dapsone 100 mg/day. At diagnosis, dermatological and neurological examination was performed on all patients. Number and characteristics of cutaneous lesions were noted. Peripheral nerves were assessed through palpation of the main nerve trunks and corresponding pain and/or thickness, when found, was noted. Sensory and voluntary muscle evaluation was done in all patients using simple techniques and Semmes–Weinstein monofilaments, by trained personnel. Patients were assessed monthly during treatment by way of
dermatoneurological examinations. The immunological tests were performed following patients’ consent for blood collection at diagnosis. Six patients (12.5%) presented reactional episodes (either reversal reaction (RR) or neuritis), 3 during multidrug treatment, and 3 during the surveillance period. Reactional lesions were biopsied to confirm clinical diagnosis of a reactional state.

**ENUMERATION OF BACILLI**

Slit smears for bacterial index (BI) determination taken from three sites (both earlobes and a cutaneous lesion) were performed at the time of diagnosis, as a routine procedure. All 48 patients were BI negative.

**ANTIGEN**

Skin tests were done with lepromin A, Carville, USA (provided by WHO). For *in vitro* tests, armadillo-derived sonicated *M. leprae* antigen was provided by Dr R. J. W. Rees (IMMLEP Bank, The National Institute of Medical Research, Mill Hill, England). Optimal antigen stimulating concentration was found to be 20 µg/ml.

**LYMPHOCYTE TRANSFORMATION TEST (LTT)**

Heparinized venous blood was collected for *in vitro* tests and peripheral blood mononuclear cells (PBMC) were isolated by Ficoll-hypaque (Pharmacia Fine Chemicals, Piscataway, NJ, USA) density centrifugation. PBMC were incubated in triplicate at 2 × 10⁵ cells/well at 37°C in 96-well U-bottom plates (Costar Corporation, Cambridge, MA, USA) in 200 µl RPMI 1640 medium (Gibco Laboratories, Grand Island, NY, USA), supplemented with 10% pooled AB serum, 100 U/ml penicillin, 100 µg/ml streptomycin and 2 mM L-glutamine (Gibco Laboratories). Cells were cultured for 5 days in the presence or absence of antigen, when supernatant was collected for determination of cytokine production. The remaining cells were pulsed with 1 µCi of ³H-thymidine for another 18 hr. Results obtained as counts per minute (cpm) are expressed as stimulation index (SI, cpm obtained from stimulated cultures/cpm obtained in control cultures).

**INTERFERON-GAMMA (IFN-γ) PRODUCTION**

The amount of IFN-γ on a 5-day culture supernatant was assayed using a commercial RIA kit (IMRX Corp., Centocor Malvern, PA), specific for the active human IFN-γ as described. Levels of IFN-γ are expressed as Units/ml (U/ml) in stimulated cultures minus U/ml obtained in control cultures. The cut-off point for positive response was 40 U/ml, which is 4 sd above mean values obtained in unstimulated cultures.

**ELISA TEST FOR PGL-1**

Serum samples from all tuberculoid patients were collected aseptically, processed under sterile conditions, and kept frozen (−20°C) until use. The presence of specific IgM antibodies anti-ND-BSA (natural disaccharide conjugated to bovine serum albumin,
which represents an immunodominant part of the phenolicglycolipid 1, PGL-1) in the serum was assayed in 21 patients as described. Plates were read in a Titertek Multiscan (Flow Laboratories), at 492 nm, and the results expressed as optical density variation (\( \Delta \text{OD} = \text{OD obtained in the antigen coated wells minus the OD obtained in the control wells} \)). A sample was considered positive when its OD exceeded by 2 sd the mean results obtained in the healthy control population. Cut-off OD value for positive response was 0.27.

SKIN TEST

Lepromin skin tests were performed by intradermal injection of 0.1 ml of the standardized crude antigen preparation (lepromin A, \(1.6 \times 10^8\) bacilli/ml, National Hansen’s Disease Center, Carville, LA, USA) in the volar surface of the left forearm. Readings were performed after 21–28 days and reactions \(\geq 5\) mm were considered positive (National Guide – Leprosy Control Program). All patients were lepromin tested before initiating specific leprosy treatment. All the injections and readings were done by the same trained person.

SKIN BIOPSIES

All patients underwent biopsies at the periphery of skin lesions before starting treatment. Biopsy specimens were processed and stained by hematoxilin and eosin, and by Wade method for acid-fast bacteria. Typical infiltrates of BT leprosy, as stated by Ridley & Jopling classification system, were found in all 48 patients, as a prerequisite for inclusion in the study. None of the patients had acid-fast bacilli identifiable in biopsy specimens.

STATISTICAL ANALYSIS

Tests were used for statistical evaluation of significance. Chi-squared test, Correlation, Regression and Somer’s Deviance were used when appropriate. Immunological tests were related to each other, and to skin lesions and nerve involvement. Standard deviation (sd) was used to express variance. Probability (\(P\)) less than 0.05 in the statistical tests was considered statistically significant.

Results

CLINICAL EVALUATION

BT patients included in the study presented characteristics clinical lesions and typical epithelioid granulomas in skin lesions, but had no detectable acid-fast bacilli in skin smears. All patients were treated with the paucibacillary WHO regime, and none of them has presented clinical diagnosis of relapse (mean of 6 years of follow-up). Among the 48 BT patients studied, 24 (50%) presented a single cutaneous lesion with well-defined borders, which were infiltrated in some patients, either anaesthetic or hypoaesthetic. The other patients presented \(\geq 2\) cutaneous lesions and at least 5 patients (10.4%) presented 10 or more lesions. Concerning the number of affected nerves at the diagnosis, 28
BT leprosy: clinical and immunological heterogeneity

Figure 1. Distribution of the 48 BT patients according to the number of (A) cutaneous lesions, and the number of (B) affected nerves.

patients (58.3%) showed only one affected peripheral nerve or none at all. Of the other 20 patients, 19 had from 2 to 9 nerves that were damaged and only one patient (2.1%) had 10 affected nerves (Figure 1). Fifteen patients (53.6%) among those with no affected nerves or only one showed a single cutaneous lesion, and 11 patients (55%) with more than one affected nerve had also more than one lesion. However, no significative correlation was observed between the number of cutaneous lesions and the number of affected nerves.

IMMUNOLOGICAL TESTS

Lepromin test

Nineteen out of 48 patients tested (39.6%) showed a positive lepromin reaction. Patients with positive lepromin tests had more skin lesions when compared to patients with negative lepromin tests (Figure 2). Although not statistically significant, it was noted that among the 19 patients positive for lepromin, 7 (36.8%) presented a single skin lesion
and 12 (63.2%) had multiple lesions. When results are expressed as the percentage of patients that responded to the lepromin test with an induration ≥ 3 mm, 91.7% of the patients showed a positive reaction. Again, no significative relationship was noted between the positivity in the lepromin test and the number of skin lesions (Table 1).

**Lymphoproliferation test (LTT)**

Among the 47 patients whose lymphocytes were submitted to LTT, 27 (57.4%) showed positive results (SI ≥ 3.0). As shown in Figure 2, no significative correlation was observed between the positive lymphoproliferative test and the number of skin lesions, but a significative relationship was shown between LTT and the extent of nerve involvement \( (P < 0.05) \). Actually, 75% of the patients unresponsive to *M. leprae* in the LTT presented only one affected peripheral nerve or none at all (Table 1). Patients with positive LTT were equally distributed in relation to the number of skin lesions and/or to the number of affected nerves. Among the 27 patients with positive LTT, 13 patients (48.1%) also had positive lepromin skin tests. In addition, 15 of the 20 patients with negative LTT were negative in the lepromin test as well. There was 59.6% agreement between these two immunological tests.
**Table 1.** Relationship among immunological tests and the number of cutaneous lesions, and among immunological tests and the number of affected nerves, in BT patients. Number of tested patients was 48 for lepromin tests; 47 for LIT; 18 for IFNγ and 21 for PGL-I

<table>
<thead>
<tr>
<th>Immunological tests</th>
<th>Number of skin lesions</th>
<th>Number of affected nerves</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>1 (+ than 1)</td>
<td>0-1 (+ than 1)</td>
</tr>
<tr>
<td>Lepromin (≥5 mm)</td>
<td>7 (36.8%)</td>
<td>10 (52.6%)</td>
</tr>
<tr>
<td>Lepromin (&lt;5 mm)</td>
<td>17 (58.6%)</td>
<td>18 (62.1%)</td>
</tr>
<tr>
<td>Lepromin (≥3 mm)</td>
<td>22 (50%)</td>
<td>25 (56.8%)</td>
</tr>
<tr>
<td>Lepromin (&lt;3 mm)</td>
<td>2 (50%)</td>
<td>3 (75%)</td>
</tr>
<tr>
<td>LTT (+)</td>
<td>15 (55.6%)</td>
<td>12 (44.4%)</td>
</tr>
<tr>
<td>LTT (−)</td>
<td>8 (40%)</td>
<td>15 (75%)</td>
</tr>
<tr>
<td>IFNγ (+)</td>
<td>4 (100%)</td>
<td>1 (25%)</td>
</tr>
<tr>
<td>IFNγ (−)</td>
<td>6 (42.9%)</td>
<td>7 (50%)</td>
</tr>
<tr>
<td>PGL-I (+)</td>
<td>1 (50%)</td>
<td>1 (50%)</td>
</tr>
<tr>
<td>PGL-I (−)</td>
<td>10 (52.6%)</td>
<td>14 (73.7%)</td>
</tr>
</tbody>
</table>

*Interferon-gamma (IFNγ) assay*

When patients’ blood ($n = 18$) was tested for the release of IFNγ into culture supernatants, only four individuals (22.2%) were found to be positive. These four patients had only one skin lesion ($P < 0.05$) (Figure 2(A)), but three of them had more than one affected nerve (Figure 2(B)). On the other hand, only 6 IFNγ negative patients (42.8%) presented a single skin leprosy lesion. Two out of the four IFNγ positive patients (50%) showed a positive lepromin reaction. In addition, eight out of 14 individuals (57.1%) with negative IFNγ were also negative in the lepromin test. There was an agreement of 55.5% between these two tests. All four patients with positive IFNγ were positive in the LTT. Of the 14 patients found to be negative in the IFNγ assay, six (42.9%) were also negative in the LTT (55.5% concordance).

*PGL-I Elisa test*

Twenty-one patients had their sera submitted to Elisa for detection of anti-PGL-1 IgM antibodies. Nineteen were negative (90.5%). No correlation was noted when compared to skin and nerve involvement. The two patients (9.5%) who showed positive results were lepromin negative, and only one had positive LTT. The low levels of anti-PGL-1 antibodies observed among these patients confirmed their tuberculoid background including those with multiple lesions.

**REACTIONAL EPISODES**

A total of 6 patients (12.5%) developed reaction. Of the four patients who developed reversal reaction (RR), two showed no nerve involvement, and two had multiple nerves affected. All patients presented more than one leprosy lesion. Two patients developed isolated neuritic episodes. Both had more than one affected nerve, but only one presented multiple cutaneous lesions. The fact that blood samples were collected from these patients during the initial examination and not during the reactional episode made the correlation between reaction and immunological tests unreliable.
Histological analysis confirmed the diagnosis of borderline–tuberculoid leprosy in all cases.

Discussion

Forty-eight borderline–tuberculoid patients were included in this study. Besides all clinical characteristics of the resistant form of the disease delineated in this study, only 50% of the patients presented single skin lesion and 58% had none or only one affected nerve. Interestingly, we have observed that patients with positive lepromin skin tests had more skin lesions, when compared to patients with negative lepromin tests. Although not statistically significant, these data emphasize the current interpretation that the lepromin test is evidence of delayed type hypersensitivity (DTH) and does not necessarily reflect protection in leprosy. On the other hand, no significative correlation was observed between the lymphoproliferative response and the number of skin lesions, but patients unresponsive to LTT had less nerve involvement, which may reflect a hypersensitivity effect. IFNγ is perhaps the key cytokine responsible for the activation of the oxidative and anti-microbial activity of human macrophages.9 The finding that all responder BT patients who released high IFNγ levels in PBMC culture supernatant had only one skin lesion ($P < 0.05$), attributes an important role to IFNγ in restraining the spread of infection. IFNγ is a key component in determining the type of effector function that eventually develops during the course of an immune response. There is considerable evidence that the presence of IFNγ in the milieu induces inhibition of the expansion of Th2-like cells. In addition, during *M. leprae* infection, a protective response is sometimes observed in the absence of granuloma formation, such as in the indeterminate form of the disease, which in some patients is supposed to be self-healing. All these data suggest that protective immune response and DTH are not the same phenomenon, in spite of being frequently simultaneous.7 However, 75% of the high responder patients in the IFNγ assay had more nerves damaged when compared to the nonresponder ones. These results have very important implications and strongly suggest that nerve damage may represent a hypersensitivity phenomenon following cell-mediated immunity against the pathogen. The potential pathological effects of IFNγ can be related to its ability to synergize with inflammatory cytokines, such as TNFα, IL-1β and others. Previous studies2 have demonstrated that IFNγ acting synergistically with inflammatory cytokines is able to induce or to increase the expression of adhesion molecules on endothelial cells and keratinocytes that affects directly the type and the amount of leukocytes attracted to the site of inflammation. Recently, a higher expression of TNFα mRNA as well as TNFα protein was demonstrated in the nerves and skin of borderline patients with RR.6 Although in some patients, absence of or low IFNγ production was detected in vitro, the immunohistochemical findings suggest that, at least in situ, levels of IFNγ are sufficient to maintain the activation state (data not shown).

Six patients presented reactional episodes (12.5%). All but one (83.3%) had more than one skin lesion. Correlation between the reactional features and the in vivo and in vitro immunological tests was not reliable. This may in part be explained by the fact
that tests were performed at the diagnosis, which, for the most part, was quite distant from the reactional episode. Neuritis following multidrug therapy seems to occur infrequently in bacteriologically negative BT patients. Only two patients (4%) presented neuritis after MDT.

In a previous report,\textsuperscript{12} it was found that seropositivity to anti-PGL-1 antibodies was associated with subsequent manifestation of RR. This association was strongest in patients who were both anti-PGL-1 antibody and lepromin positive. However, to our way of thinking, no correlation was noted between levels of anti-PGL-1 antibodies and development of reaction. The two patients who were anti-PGL-1 antibody seropositive were lepromin negative and were not included among the six individuals who developed reaction during the study.

The present study confirms previous reports indicating the role of IFN\textgreek{g} in mediating resistance to mycobacterial infection.\textsuperscript{4,5} However, other mechanisms might be involved in restricting bacterial growth in borderline–tuberculoid patients.

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