

Comments on the Chemotherapy of Leprosy as Influenced by Present Knowledge of *Mycobacterium leprae**

S. R. PATTYN

*Professor of Bacteriology,
Instituut voor Tropische Geneeskunde Prins Leopold,
Antwerp, Belgium*

The chemotherapy of leprosy can now be based on firm laboratory knowledge about the responsible agent, *Myco. leprae*, and the general principles applied in the management of tuberculosis. First-line and second-line drugs may be distinguished. At present there is no need to change the existing dosage of 600 mg of dapsone (DDS) per week for mass campaigns.

Efforts should be made to find new treatment schedules leading to increased supervision by the application of intermittent therapy. In lepromatous cases, this intermittent schedule will probably have to be preceded by a preliminary course of continued therapy. It is possible that the use of rifampicin will appreciably shorten the period of treatment.

Introduction

Although an increasing number of new drugs have been introduced in the therapy of leprosy during the last three decades, their choice has been largely empirical, due to lack of knowledge about the responsible organism, *Mycobacterium leprae*, and the pathogenesis of the disease.

In the last 10 years, however, as a result of laboratory investigations, our knowledge and understanding of leprosy have increased more than in the 87 years between 1873 (when *Myco. leprae* was discovered) and 1960. In the latter year Shepard (1960a, b, 1962) demonstrated, using precisely defined conditions, that *Myco. leprae* multiplied in the footpads of mice. The second important discovery—the cellular type of immunity involved in leprosy—was made by Rees (Rees, 1966; Rees *et al.*, 1967) who showed that a generalized infection resulted when thymectomized and irradiated mice were inoculated with *Myco. leprae*. Several authors have since shown that a cell-mediated type of immunity is involved in the human disease (Dierks and Shepard, 1968; Shepard, 1968; Turk and Waters, 1969).

These discoveries revealed new features about *Myco. leprae*, such as its long generation time (Shepard and McRae, 1965), the interpretation of irregularly stained organisms (Rees *et al.*, 1960; Rees and Valentine, 1962; Shepard and McRae, 1965) and their quantitative assessment, using the Morphological Index

* Accepted for publication, 19 July, 1972.

(M.I.). Moreover, a more accurate histo-pathological classification of the various forms of human leprosy was evolved by Ridley and Jopling (1966) by including immunological features of the disease. Finally, for the first time a rational approach to the evaluation of antileprosy drugs in mice and man was developed by including all these new parameters.

In mice, antileprosy drugs can be assessed in three ways: (1) by the continuous method (Shepard and Chang, 1962; Pattyn and Rooyackers, 1965; Rees, 1967a; Gaugas, 1967); (2) by the kinetic method; and (3) by treatment of an established infection (Shepard and Chang, 1967). In man, the activity of antileprosy drugs can be monitored by the regular inoculation into mice of *Myc. leprae* obtained from human biopsies taken during treatment (Shepard *et al.*, 1968, 1969; Levy *et al.*, 1969; Rees *et al.*, 1970). These methods have revealed: (a) the time necessary for drugs to kill *Myc. leprae* (Shepard *et al.*, 1968; Rees *et al.*, 1970); (b) the existence of drug-resistant strains of *Myc. leprae* (Pettit *et al.*, 1964, 1966a, b; Rees, 1967b; Pearson *et al.*, 1968; Shepard *et al.*, 1969); (c) the correlation existing between irregularly-stained forms of the bacilli and their inability to multiply, proving that non-solid *Myc. leprae* are dead.

There has been some divergence of opinion, and even scepticism, about the significance of the percentage of non-solid forms of the bacilli (Convit *et al.*, 1970). Indeed, all morphological assessment is subjective. Therefore standard techniques in fixation, staining, observation and scoring must be used for determining the M.I. Perhaps some future histochemical method will supersede the counting of solidly staining (living) bacilli. At present the morphological criterion can only be checked by mouse inoculation, a technique that is entirely reliable (Levy *et al.*, 1970).

This newly acquired knowledge about *Myc. leprae*, together with advances made in the management of tuberculosis, another chronic infectious disease, now provide a rational basis for studying leprosy therapy. In the chemotherapy of tuberculosis, experience has shown that successful or unsuccessful therapy, relapse, or drug resistance, is determined by the biological behaviour of the causative organism and the pharmacokinetics and pharmacology of the drugs used. Studies of all these aspects, together with controlled clinical trials, have led step by step to the development of completely successful therapeutic regimens, of which the latest is based on intermittent therapy (after an initial continuous period) allowing complete supervision (Stradling and Poole, 1963, 1970). While the importance of maintaining the prescribed therapy throughout for the successful treatment of chronic infections has been particularly appreciated and employed in tuberculosis, similar considerations seem to have been less rigorously applied to the treatment of leprosy.

There are three factors to be taken into account in planning a rational approach to drug therapy of chronic infections in general, and leprosy in particular. These are discussed below under the headings: the bacillus, the host, and the drug.

The Three Interacting Factors in Drug Therapy

THE BACILLUS

For a correct interpretation of events during the therapy of leprosy, the notion that non-solidly staining bacilli are incapable of multiplication is of great importance. It is now certain that, while a population of *Myc. leprae* may be killed by treatment within a few weeks or months, it may take 5 years or more

for the bacilli to disappear from the skin or nasal mucosa. Thus the M.I. (if necessary checked by inoculation of bacilli from treated patients into mice) is of primary importance for following the effect of treatment (Shepard *et al.*, 1968, 1969; Levy *et al.*, 1969; Rees *et al.*, 1970).

Since multiplication of *Myc. leprae* in the mouse footpad is limited to inocula of 10^4 to 5×10^4 bacilli and the minimal infectious dose is between 3 and 10 organisms (Shepard and McRae, 1965), the sensitivity of the technique is of the order of 1/1000 living bacilli.

The total bacillary population of a patient at the start of treatment not only determines the severity of the disease but also the number of pre-treatment drug-resistant bacilli. Indeed, in every bacterial population there exist resistant mutants to any drug, but these spontaneous mutants ("pre-treatment resisters") have an advantage over the larger, sensitive part of the population only when exposed to the drug. For most pathogenic organisms, such as *Myc. tuberculosis*, the frequency and level of drug-resistant mutants can be measured *in vitro* by counting the number of colonies in media containing known concentrations of the drug and seeded with known numbers of organisms (Canetti *et al.*, 1961). In general, the frequency of such pre-treatment resistant organisms is of the order of 1 in 10^6 to 10^8 . However, although similar *in vitro* determinations are not possible for *Myc. leprae*, drug resistance can be expected to arise in multi-bacillary types of leprosy (LL-BB) with between 10^{10} and 10^{12} organisms; but this is unlikely in the paucibacillary types (BT-TT).

The level of drug resistance of the "pre-treatment resisters" is also unknown for *Myc. leprae*. However, the large population of bacilli obtained in the immunologically deficient mouse might be sufficient to determine the level of drug resistance of these mutants. From available clinical data it appears that the degree of drug resistance is variable (Rees, 1967a, b; Pearson *et al.*, 1968).

THE IMMUNOLOGICAL COMPETENCE OF THE HOST

In the chemotherapy of most infectious diseases, the drug used accounts for the killing of the majority of the bacteria, while the defence mechanisms of the host deal with the remainder. In tuberculosis (Canetti, 1965), these residual organisms (drug-sensitive or drug-resistant) may persist, sometimes apparently in a dormant state, for a long time. In this situation, these organisms may begin to multiply again at any moment after treatment has ceased, and the patient relapses with a drug-sensitive or a drug-resistant infection.

In the patient with the lepromatous type of leprosy, the host has little or no capacity to destroy bacilli, and therefore *all* persisting and viable organisms will multiply again when treatment is stopped. This accounts for the high relapse rates reported in lepromatous leprosy: 12% in India (Vellut, 1968) and 25% in Latin America (Bechelli *et al.*, 1970), and Hemerijckx (personal communication) recommends that lepromatous patients should be treated for life.

THE DRUG

(a) *Minimal Inhibitory Concentration (M.I.C.)*. The M.I.C. of many drugs against *Myc. leprae* can now be determined by the mouse footpad technique. The results (Table 1) show that there is no correlation between the antibacterial activity of drugs against *Myc. tuberculosis* or *Myc. lepraemurium* and *Myc. leprae*.

(b) *Serum concentration and half-life of the drug*. This may show considerable individual variation, and therefore the lowest figures known are given (Table 1).

TABLE 1

M.I.C. for several antileprosy drugs compared with serum concentrations obtained after current dosage

Drug	M.I.C. ($\mu\text{g/ml}$)	Dosage	Frequency	Serum concentration ($\mu\text{g/ml}$)	X
DDS	0.02-0.002	100	Daily	2	100
DADDS	id.	225 mg	1/77 days	0.06	3
Sulfadimethoxine	20	1.5 g	Daily	150-300	4-15
Sulfadoxine	20	1.5 g	1/week	150-300	4-15
Sulfamethoxy-pyridazine	0-35?	750 mg	1/2 days	30-40	4-15
Clofazimine	Current dosage is 100-200 mg per day Minimal effective dosage in man calculated to correspond to 7 mg per day				
Thiambutosine	?	3 g repository	Daily	?	
Ethionamide	25	2 g 250 mg	1/week Daily	20-30	1

Genetic polymorphism in man has recently been described for the acetylation of dapsone (=inactivation) comparable to that shown for isoniazid (INH) (Gelber *et al.*, 1969) and may be of practical importance when intermittent or low dosage DDS is administered, as was the case in intermittent treatment of tuberculosis with INH (Tuberculosis Chemotherapy Centre, Madras, 1970).

Another important point is the rate at which drugs kill *Myc. leprae*, for which the following results are available:

DDS: 90 days (Shepard *et al.*, 1967, 1968)

Clofazimine (including DDS-resistant strains): more than 105 days (Levy *et al.*, 1969)

Rifampicin: 3 to 24 days (Rees *et al.*, 1970).

Thus, if a patient is treated with a drug having a high therapeutic activity, and he does not improve, or after some period of improvement there is clinical and bacteriological deterioration, there are only two explanations: either the infecting organism was or has become drug-resistant, or the patient failed to take or absorb the prescribed treatment (Poole and Stradling, 1960). This leads to the rather astonishing conclusion that in many cases the only advantage to be gained from determining the drug sensitivity of bacilli during treatment may be to demonstrate that the patient is taking the drug (McDermott, cited in Canetti, 1965). The persistent failure of leprosy patients to take their drugs regularly has been particularly well investigated by Pettit *et al.*, (1966); some patients with lepromatous leprosy still active after more than 10 years' residence and treatment in a leprosarium, improved when the treatment was adequately supervised.

The Possible Application of Principles Used in the Chemotherapy of Tuberculosis

In the chemotherapy of tuberculosis a distinction is made between first-line drugs (low M.I.C., sustained serum levels many times higher than the M.I.C., minimum toxicity, and low incidence of pre-treatment resistant organisms) and

second-line drugs (high M.I.C. and relatively low serum levels of shorter duration, resulting in more frequent resistance and/or higher toxicity). Treatment of multibacillary cases of tuberculosis with cavitation should always be initiated with 2 (preferably 3) major drugs in order to diminish the risk of the emergence of drug resistance, the frequency of double or triple drug resistance being of the order of 10^{-12} to 10^{-18} . After a preliminary period of continuous double or triple treatment, reducing bacterial numbers drastically and thus the possibility of drug resistance, long-term monotherapy can safely be given (Canetti, 1968). To ensure that the drugs are being taken treatment should be supervised, and for practical purposes this is easier to achieve by an intermittent, rather than a daily, regimen (Poole *et al.*, 1960; Stradling *et al.*, 1963, 1970). Moreover, intermittent therapy has a theoretical advantage, because contact between the micro-organism and the drug is discontinuous. Thus the surviving organisms do not multiply again immediately, but only after a lag phase. This phenomenon has been demonstrated experimentally both *in vitro* and *in vivo* (Dickinson, 1968; Canetti, 1968). Experimental studies with *Myc. leprae* on intermittent therapy in mice (Rees, 1965, 1967), using Shepard's kinetic methods (Levy, 1970), suggest that this phenomenon applies equally to leprosy.

Treatment of Leprosy

DAPSONE

The activity of substituted sulphones is entirely due to dapsone (4,4-diaminodiphenylsulphone, DDS), either liberated after administration or occurring as an impurity in some preparations (Shepard, 1969a; Shepard *et al.*, 1969). The M.I.C. of dapsone for *Myc. leprae* is of the order of $0.02 \mu\text{g}$ per ml serum, as determined by means of the mouse footpad model (Shepard *et al.*, 1966; Shepard, 1967b; Rees, 1967). Because the metabolism of dapsone is similar in mouse and in man, the results obtained in the mouse are directly applicable to man. Experiments in man have shown this to be the case. Thus, the administration of 1 mg of dapsone daily, or the intramuscular injection of 225 mg of acedapsone (acetyl-diaminodiphenylsulphone, DADDS) every 77 days, produced respectively serum levels of 0.02 and $0.06 \mu\text{g}$ per ml of dapsone and proved effective in the treatment of leprosy in man (Shepard *et al.*, 1968; Waters *et al.*, 1968).

When dapsone was introduced for the treatment of leprosy 30 years ago the dose chosen was based on the maximum amount tolerable (100 mg daily), because then the mouse model was not available (Lowe, 1954a, b). It is now proven that this dose produces serum levels of dapsone 100- to 300-fold the M.I.C. for *Myc. leprae*. Present results in mouse and man suggest that therapeutically the dose of dapsone could be reduced, but lower doses might increase the incidence of drug resistance.

The observations of Pettit *et al.* (1966), Shepard *et al.* (1969), and Browne (1969) have shown that DDS resistance can occur in some lepromatous patients on standard, irregular, or smaller (50 mg twice weekly) doses of dapsone. Resistance may well arise if acedapsone, 225 mg every 77 days, is used on a large scale. However, because of the long generation time of *Myc. leprae*, it may take 3 or more years before dapsone-resistant mutants give rise to clinical evidence of relapse. The rarity of dapsone resistance noted during the 30 years of standard dapsone monotherapy is probably because there are very few high-resistant mutants capable of growing at the high concentrations of dapsone obtained.

For the treatment of tuberculoid leprosy, in which there is a small bacterial population and therefore fewer resistant mutants than in patients with lepromatous leprosy, lower doses of dapsone (1/10th of the standard dose) could be adequate. However, because of the low cost and low toxicity of dapsone in the standard dosage, smaller doses would have no practical advantages, whereas one standard dose for all types of leprosy would be advantageous.

For the treatment of lepromatous leprosy, lower doses of dapsone are much more likely to give rise to resistance, particularly if taken irregularly. Therefore lower doses of dapsone in lepromatous leprosy could be justified only when preceded by an initial period of standard doses (100 mg) intended to diminish the bacterial load considerably without danger of selection of resistant organisms. Lower doses are also justified when they are attained by intermittent therapy and therefore increase the possibility of supervised treatment. It is possible that, in the past, standard doses of dapsone (100 mg daily) were curative even in those patients who, after an initial period of regular treatment, became irregular, but took sufficient dapsone to produce active drug levels in the serum. This possibility is based on the fact that serum levels of dapsone obtained from standard treatment are 100- to 300-fold greater than the M.I.C. of dapsone for *Mycobacterium leprae*.

Thus it would seem important to initiate controlled clinical trials in patients with lepromatous leprosy, with 4 to 6 months high dose (100 mg) continuous dapsone therapy, followed by supervised intermittent therapy, for example, 25 mg dapsone twice weekly *per os*, or once weekly long-acting sulphonamides, or injections of acedapsone (DADDS) every 77 days (Shepard *et al.*, 1968).

As has already been mentioned, relapses can be due either to drug-sensitive persisters multiplying after therapy has been stopped, or to the emergence of drug-resistant mutants. To determine the cause of such relapses, the sensitivity of the organisms can be tested in the mouse. However, the patient who has relapsed should also be put on a supervised test-period for 3 months on full-dose dapsone and any change in the morphological index used as an indirect method to detect a dapsone-sensitive or dapsone-resistant infection. This clinical method is more rapid than the mouse test. If it is confirmed that the histoid type of leprosy is always associated with dapsone-resistant organisms (Rodriguez, 1969), histopathological examination would rapidly resolve the question.

LONG-ACTING SULPHONAMIDES

Sulphones and sulphonamides both interfere with folate metabolism (Shepard, 1967) and cross-resistance between the two types of drug exists (Adams and Waters, 1966). Some sulphonamides were tried in the treatment of leprosy before the sulphone era (Schneider *et al.*, 1959); they were reintroduced as a result of a chance observation (Schneider *et al.*, 1959). The dose of sulphonamides for leprosy was chosen empirically and resulted in serum levels of 30 to 40 μg per ml (Schneider *et al.*, 1959; Languillon, 1964; Languillon and Carayon, 1969). Ellard *et al.* (1970) found the M.I.C. against *Mycobacterium leprae* in the mouse for sulphadimethoxine and sulphadoxine to be 20 and 35 μg per ml respectively, and the calculated serum levels attained in man with these two drugs to be 150 and 300 μg per ml respectively. Since these serum levels are only 4 to 15 times higher than the M.I.C., Ellard *et al.* (1970) warned that sulphonamide resistance and, because of cross-resistance, sulphone resistance might result from treating patients with lepromatous leprosy with these drugs. Sulphonamide resistance has been

observed in man (Merklen *et al.*, 1968) and most authors (Schneider *et al.*, 1959; Litalien *et al.*, 1961; Languillon, 1964) consider sulphonamide treatment to give better results in tuberculoid than in lepromatous leprosy. The laboratory findings indicate that the use of long-acting sulphonamides should be limited to the treatment of tuberculoid leprosy, and, in lepromatous leprosy, as intermittent therapy following initial treatment with a bactericidal drug.

CLOFAZIMINE (B663)

Current dosage, again empirically determined, has been between 100 and 200 mg per day. Since this drug has different affinities for different tissues, dosage cannot be based on the M.I.C. and serum levels. However, Shepard (1969b) calculated the minimal effective dosage in man to be 7 mg per day. Studies by Levy *et al.* (1969) on the killing rate of *Myc. leprae* by clofazimine showed it to be somewhat slower than that of dapsone. Clofazimine is an important drug because of its low toxicity, high activity, and absence of cross-resistance with the sulphones or sulphonamides. Its great disadvantage is the skin pigmentation it induces, which is unacceptable to many patients. Some patients, however, appreciate this discoloration (Renders, 1968), and on the other hand it confirms that the patient is taking the drug.

Where skin pigmentation is a major obstacle, a dose of 200 mg per week (Waters *et al.*, 1968) would probably be adequate for patients with tuberculoid leprosy or as a secondary drug for patients with lepromatous leprosy following initial therapy with a bactericidal drug. Controlled trials along these lines seem indicated.

RIFAMPICIN

The M.I.C. of rifampicin for *Myc. leprae* has not yet been determined. However, its extraordinarily rapid killing effect on *Myc. leprae* is well known. Thus, it has been observed both in man (Rees *et al.*, 1970; Leiker *et al.*, 1970, and our own unpublished observations) and in the mouse (Shepard, 1971) that 2 days' treatment with rifampicin is as effective as 2 to 3 months of dapsone. Grumbach *et al.* (1969) considered that, for tuberculosis, rifampicin, in spite of its price, would be advantageous by shortening the duration of therapy. Future studies will determine whether a relatively short course of rifampicin will cure tuberculoid leprosy. In lepromatous leprosy, rifampicin might with advantage be given initially and continuously for a relatively short period, say 3 months, followed by intermittent treatment with either acedapsone, a sulphonamide, or clofazimine. Laboratory controlled treatment schemes along these lines are recommended.

THIAMBUTOSINE (DPT)–THIACETAZONE (TB1)–ETHIONAMIDE

The appearance of resistance to these drugs after 1 to 2 years of treatment is a regular phenomenon (Lowe, 1954b; Davey, 1955, 1960) and many resistant strains have been isolated in mice (Shepard, 1969; Rees, 1967b; Pattyn *et al.*, 1965). This must be due to the low serum levels attained with the current dosages and these cannot be increased because of the high risk of toxic side-effects (Cochrane and Davey, 1964). Thus thiambutosine is only a "second line drug" for leprosy; it took many years to reach this conclusion, despite the fact that thiambutosine was the second antileprosy drug discovered. The delay was due to the lack of suitable laboratory techniques which are now available. Ethionamide is also a "second line drug" that develops cross-resistance with related drugs (Floch

et al., 1966). The M.I.C. of ethionamide for *Myc. leprae* in the mouse is 25 μg per ml (Shepard, 1969b). This value is very close to the serum levels calculated to attain 20 to 30 μg per ml when given at a dosage of 250 mg daily, as was done by Floch *et al.* (1966). These authors found ethionamide to be active, but the duration of the trial was manifestly too short (4 to 18 months). Moreover, with the additional knowledge that ethionamide is rapidly excreted, that inhibition of the growth of *Myc. leprae* is of short duration (Shepard, 1969), and that the dosage cannot be increased for long-term treatment (because of toxicity), it is clear that ethionamide will remain at best a second-line drug.

Treatment of Patients with Drug-resistant Organisms

The occurrence of drug resistance to thiambutosine and dapsone is well-established. Because thiambutosine is a second-line drug in leprosy, it should not be administered alone at the start of treatment of patients with multibacillary disease. Cases resistant only to thiambutosine are not a therapeutic problem, since first-line drugs will still be effective. However, patients who are resistant to dapsone need special attention for two reasons. The first is the potential danger of spread of dapsone-resistant strains of *Myc. leprae* in the community, although nothing is yet known about their infectiousness for man. For the present, dapsone-resistant strains should be considered infectious for man; for the mouse, they are as infectious as dapsone-sensitive strains. The second reason is psychological. If dapsone resistance appears in a patient for whom high dosage dapsone (500 to 600 mg per week) has been prescribed, then he has probably taken the drug irregularly and not in the way prescribed. He may well be irregular with any new therapy and therefore will relapse again. Such patients present individual and community problems. Moreover, their apparently "uncured leprosy" may be quoted or used by others to prove, wrongly, that "leprosy is incurable".

Rifampicin and clofazimine are fully effective against dapsone-resistant strains of *Myc. leprae* in the mouse, and therefore are the drugs of choice for the treatment of patients with dapsone-resistant bacilli. Determination of the level of dapsone-resistance is only useful for identifying patients with intermediate levels of resistance, still capable of responding to full doses of dapsone (100 mg per day). However, for such patients dapsone should be given only in combination with one or two other antileprosy drugs.

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Note added in proof. Since the submission of this manuscript, Holmes and Hilson (*J. med. Microbiol.*, 1972, **5**, 251) have published the results of rifampicin treatment of the experimental infection of mice by *Myc. leprae*. The results indicate that rifampicin is very rapidly bactericidal and that the MIC is around 0.3 µg/ml whereas the serum concentration attained in man during rifampicin treatment with 600 mg/day "fluctuates through the day from about 15 µg/ml to 0.5 µg/ml". The bactericidal effect of rifampicin on *Myc. leprae* was confirmed in the mouse model.